



MRC PPU
**Reagents
and Services**

ANTIBODY TESTING RESULTS
Standard Reporting Template

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Antibody Name: Anti-IBV-NS1

Full Antigen Name: NS1 (non-structural protein 1) protein of influenza B virus (IBV) strain B/F1orida/04/2006

Antigen Species: Virus

Sheep Number: DA280

Bleeds Tested: 1st, 2nd, 3rd, 4th, 5th

Recommended Bleed: All bleeds suitable for applications listed.

Immunoblotting:

Method

For infections, MDCK cells were mock infected, or infected with IBV strain B/Florida/04/2006 at MOI of 0.01 PFU/cell.

For transfections, 293T cells were either mock transfected, or transfected with IBV pPolI:NS (a reverse genetics plasmid that encodes the NS1 and NEP proteins of B/Florida/04/2006 as vRNA) and the four pCAGGS helper plasmids from a 12-plasmid reverse genetics system (which encode the polymerase proteins and nucleoprotein, which together transcribe vRNA into mRNA). Transfections were performed using Lipofectamine 2000 (Thermo Fisher) with cell density and plasmid mass as recommended in the manufacturer's protocol.

Cells were lysed in Laemmli buffer at 48 h post-infection or 48 h post-transfection, separated by electrophoresis on precast Any kD polyacrylamide gels (Bio-Rad) and transferred to nitrocellulose membranes.

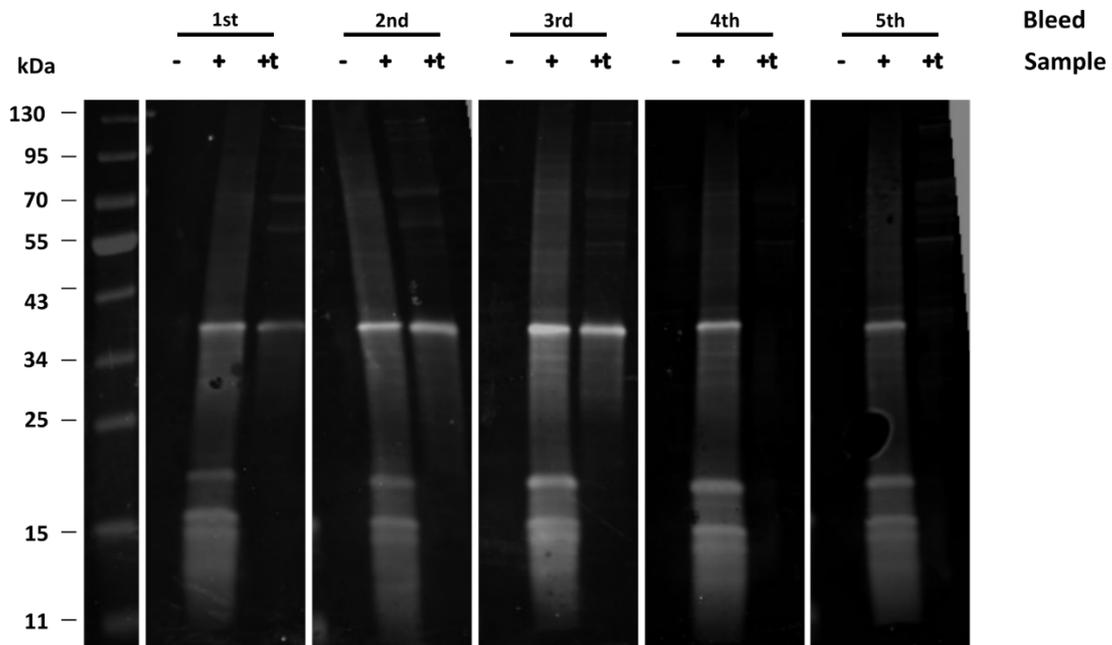
Membranes were blocked with 5 % milk in PBS-T (phosphate buffered saline with 0.1 % Tween 20) overnight at 4 °C, rinsed in PBS-T then probed with primary antibodies at 5 µg/ml in PBS-T for 1 h at room temperature. Membranes were washed ×3 in PBS-T followed by further incubation with anti-sheep IR680 or IR800 (Thermo Fisher) at 1 in 10,000 in PBS-T, for 1 h at room temperature. Membranes were scanned with a Licor Odyssey CLx Infrared imaging system.

Results

IBV NS1 = ~30 kDa. Ladder is the Page Ruler Prestained NIR Protein Ladder (Thermo Fisher).

+ infected cell lysate

+t transfected cell lysate



Signal was detected for infected cell lysates for all bleeds and for transfected cell lysates for bleeds 1 – 3, at 5 µg/ml. However, it should be noted that, although the band was not seen in the mock, it migrated more slowly than predicted.

Recommendation

All antisera can be used at 5 µg/ml for western blot with infected cell lysates, and bleeds 1 – 3 can be used with transfected cell lysates.

Immunofluorescence:

Method

MDCK and 293T cells were seeded at 2×10^5 cells/well in 24 well plates containing 13 mm coverslips.

For 293T cells, coverslips were first coated with poly-D-Lysine to promote adhesion.

The day after seeding, MDCK cells were infected with IBV at an MOI of 2 PFU/cell and 293T cells were transfected as above using Lipofectamine 2000, following the manufacturer's protocol.

Cells were fixed at 24 h post-infection or 48 h post-transfection in 4 % formaldehyde in PBS, then permeabilised using 0.2 % Triton-X100 in PBS/2 % fetal bovine serum (FBS) before blocking for up to 1 h in PBS/2 % FBS.

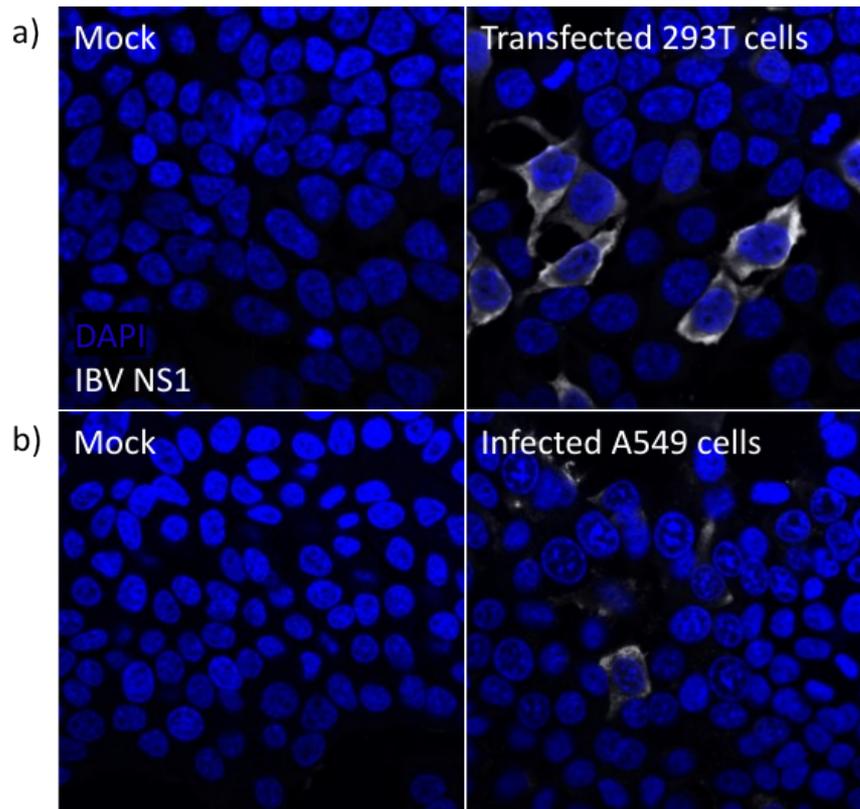
All antibody dilutions were carried out in PBS/2 % FBS. Cells were incubated with primary antibodies at 1:50 for 1 h at room temperature, washed 3 times in PBS/2 % FBS then probed with donkey anti-sheep IgG (H+L) Cross-Adsorbed Secondary antibody, AlexaFluor 647(Invitrogen) at 1:500 and DAPI at 1:1000 for 45 minutes at room temperature.

Cells were washed 3 times in PBS/2 % FBS and once in PBS, excess moisture was removed before mounting coverslips on slides using ProLong Gold Antifade mounting agent (Thermo Fisher).

Mounting agent was allowed to harden overnight before observation using a Zeiss 710 confocal microscope.

Results

A specific signal was detected for all bleeds in both infection and transfection. Images show 3rd bleed antibody at 24 h post-infection and 4th bleed antibody at 48 h post-transfection; similar results were observed for the other bleeds.



- a) Signal detected using all bleeds at 1 in 50 dilution in NS transfected cells.
- b) Signal detected using all bleeds at 1 in 50 dilution in IBV infected cells.

Recommendation

All 5 bleeds are suitable for use in IF of both infected and transfected cells at a 1 in 50 dilution.

PUBLICATIONS:

None to date.